

Role of Interleukin-1 Beta in Orthodontics

Dr. S. Lalithapriya¹, Dr. K. Rajasigamani², Dr. V. Bhaskar³

¹Ph.D Research scholar, ²Dean & HOD, ³Professor,
Department of Orthodontics & Dentofacial Orthopaedics, Rajah Muthiah Dental College & Hospital, Annamalai
University, Chidambaram, Tamil Nadu, India. Pin-608002

Corresponding Author: Dr. S. Lalithapriya

ABSTRACT

The core of orthodontic treatment philosophy was to move the teeth through the bone to a favourable position thereby obtaining an aesthetically and functionally stable occlusion. The mechanical force was the therapeutic tool used by an orthodontist during orthodontic treatment. An efficient orthodontic tooth movement can be accomplished with the appropriate use of mechanics and adequate knowledge about the biological response. Interleukin-1beta, a member of cytokine family was one among the various molecules that got activated and released in response to the applied mechanical force in tissues and cells surrounding the tooth structure and plays a key role in initiating an aseptic inflammatory response which was not only a prerequisite for bone remodelling and tooth movement but also had an effect on other associated phenomenon such as pain and resorption during orthodontic tooth movement. This paper contains a brief description on the significance of Interleukin-1beta in orthodontics and an attempt had been made to understand the cross talk between the pathways leading to bone remodelling, pain, tooth movement, and root resorption as they all have inflammation as a common thread.

Key Words: IL-1beta, Inflammation, OTM, EARR, pain, bone remodelling.

INTRODUCTION

The transduction of mechanical stimuli from an orthodontic appliance to the cells in periodontal tissue triggers a biological response that facilitates the remodelling of alveolar bone and periodontal ligament (PDL) resulting in orthodontic tooth movement (OTM). This biological response is an inflammatory reaction but it does not represent a pathological condition hence it is considered as an aseptic inflammation. The findings of DAMP system by Chen and Nunez [1] further supported the concept of aseptic inflammation. Orthodontic force induced inflammatory mediators have been identified and these were listed out by Yamaguchi and Garlet. [2]

Cytokines are one among them and it grabbed its attention in the field of orthodontics due to its many characteristics features. Interleukin - 1 beta (IL-1 beta) is an extensively studied cytokine expressed in gingival crevicular fluid (GCF) in orthodontic literature. It is a potent pro inflammatory and bone resorptive marker. The aim of this review is to give an insight about Interleukin -1 beta and its role in orthodontic tooth movement and its associated phenomenon.

History and background

Cytokines-Interleukins:

Cytokines are pleiotropic, they can be redundant, they synergize and antagonize each other, create cascade effect by stimulating the production of other

cytokines. They have influence over the expression of cytokine receptors. Cytokines can be grouped into interleukins (IL), tumour necrosis factors (TNF), chemokines, interferons (IFN), and growth factors. Cytokines involved in a variety of immune and inflammatory activities. Depending on their action the cytokines may be pro inflammatory (increase the inflammatory response), anti-inflammatory (decreases the inflammatory response), Macrophage activating cytokines, B cell activating cytokines, T cell activating cytokines, Eosinophil and/ or mast cell activating cytokines, inhibition of virus replication. Interleukins that are associated with bone remodelling include IL-1 alpha, IL-1 beta, IL-2, IL-3, IL-4, IL-6, IL-8, IL-13. [3] Among these, IL-1 beta is the earliest marker identified for bone resorption. [4] It is one of the seven molecules (IL-1 alpha, IL-1 beta, IL-18, IL-33, IL-36 alpha, IL-36 beta, and IL-36 gamma) with agonist activity within the IL-1 family ligands which constitutes in addition to the seven the molecules, three receptor antagonists (IL-1Ra, IL-36Ra, and IL-38), an anti-inflammatory cytokine (IL-37) and there are eleven molecules included in the IL-1R family. [5]

Discovery of IL-1 beta:

The discovery of IL-1 beta began in 1948 when Paul Beeson reported in his investigation on fever, the release of an endotoxin-free protein material known as endogenous pyrogen from a rabbit. [6] After forty years this material was further explored by Patrick Murphy and Barry Wood. [7] Bodel et al [8] in 1977 found in their study that this material was not pre-formed but it was released by human peripheral blood following *de novo* synthesis. Later in 1980 Bodel et al [9] found this material which came to be known as IL-1 beta in human white blood cells, as well as Hodgkin's and lymphoma cells.

Location, Synthesis, activation and signalling mechanism:

It is located on chromosome 2 between IL-1alpha and IL1-Rn, 40 to 110kb from IL-1alpha. The synthesis and activation of IL-1 beta are regulated by various mechanisms. The expression of IL-1 beta in the cytoplasm is brought about by inflammatory signals and it is not constitutively expressed. [10] It is expressed in pro-form and it is converted into an active form by caspase-1, a type of cysteine protease while caspase -1 gets activated by inflammasome, a large cytoplasmic multiprotein complex.

Interleukin 1 beta in orthodontics

Interleukin 1 beta and pain:

It was well established in the literature about the effect of Cytokines in pain. [11] Orthodontic pain commonly accompanies OTM in almost all the treatment stages of orthodontic treatment. Orthodontic pain is inflammatory in nature resulted from the force induced vascular occlusion in the periodontium. The response includes the interaction of vascular, cellular and chemical events. [12] Interleukin-1 beta one of the chemical mediators released during tooth movement encourages secretion of pain producing substance and they have a key role in activating or sensitizing nociceptors in the periodontium around teeth and orthodontic pain is mediated by these nociceptive fibres in the periodontium. [13-16] The nociceptive stimuli picked up by the nerve fibres in periodontium are transmitted to Central nervous system (CNS) via three-order neurons namely, trigeminal ganglia, the trigeminal nucleus caudalis and the ventroposterior nucleus. [12] As pain and immune system are interrelated it often becomes difficult to say whether the reduction in production of pro inflammatory cytokines reduce the pain or blocking nociception results in the lesser formation of proinflammatory cytokines. [17] Stimulation of inflammation mediators results in the release of neuropeptides which in turn

enhance the inflammatory reaction by again stimulating the inflammatory factors which trigger the release of more neuropeptides thus creating a vicious cycle. Neurogenic inflammation is an inflammatory reaction enhanced by the neuropeptides. The expression of neuropeptides in periodontium and dental pulp during orthodontic tooth movement indicates its role in neurogenic pain during orthodontic treatment. [18,19] studies have found that the immune and nervous system are linked by cytokines and they may play a role in pain and hyperalgesia. Among these cytokines, IL-1 activates or sensitise the nociceptor fibres by directly or indirectly by a complex signalling cascade. [20-23] Studies have been carried out to find out the correlation between the levels of interleukin – 1 beta in GCF and pain experienced during orthodontic treatment. [15,24-26] In spite of the increased number of research in the field of pain utilizing orthodontic tooth movement as a model to explore the complex mechanisms of neuronal involvement, missing links still exist in the pathway of pain during orthodontic tooth movement.

Levels of Interleukin 1 beta in oral fluids and tooth movement/force or stress applied:

During the last decades, various studies have been extensively done to understand the changes in the periodontium associated with OTM. The recent advance in cellular and molecular biology played a way to investigate the role of different PDL cells in bone remodelling during orthodontic tooth movement. [27-30] Studying the mediators in oral fluid (both saliva /GCF) will help us to understand the changes taking place in the periodontium as these mediators reflect the microenvironment where the force being transferred, in addition, it supports us in assessing and improving the orthodontic tooth movement. [31] As a result of force application, an adequate amount of IL-1 beta produced in the periodontal ligament diffuse into the GCF and studies have identified IL-1 beta

has a biomarker of orthodontic tooth movement. [32,33]

Among the various regulatory proteins detected in gingival crevicular fluid, cytokines is a protein of interest as they play an important role in cell signalling and bone remodelling. [34-36] Various clinical studies have been carried out to study the up and down regulation of these cytokines from GCF and its role in orthodontic tooth movement. [25,37,38]

Since IL-1 beta is produced under mechanical stress, studies have been done to discover the relationship between force application (intensity, duration) and production of IL-1 beta in GCF. Production of IL-1 beta reaches its peak levels differently in different studies (24 hr/ 4hr/8hr/3 days/7days/21 days/2 months/ 3 months/6months). [15,24,32,37,39-43] Influence of Type and amount of force on the expression levels of IL-1 beta had also been considered by certain authors. Lee et al [44] studied and compared the influence of continuous and interrupted force on the expression levels of IL-1 beta. They stated that for continuous force the level reached its peak at 24hr while for interrupted force it reaches after the first activation of the appliance. While Luppapanornlarp et al [15] compared 150g and 50g force and evaluated the levels of IL-1 beta and pain in response to these force. He concluded in his study that there is a rise in the level of IL-1 beta expression and pain in higher force while 50g of force could produce similar tooth movement with less pain. Certain studies did not concentrate on the peak value but they correlated the fluctuated levels of IL-1 beta: IL-1 RA ratio (activity index) with the velocity of tooth movement when various stresses applied to the teeth. [40,45-47] Being an inexpensive and non-invasive method, whole saliva samples can be considered as an alternative to GCF for analysing markers that reflects the periodontal environment [48] and the mediators can be detected as they are washed out into saliva from the gingival crevice. Noor Saadi and Ghaib [49] in their study estimated IL-1 beta in saliva and

find an association between the force applied and IL-1 beta during tooth movement. Luppapornlarp and Iida^[50] in their review stated that the most of the studies reporting the association between IL-1beta or receptors in GCF with tooth movement or pain or force are arriving at a conclusion that there is an instant release of IL-1 beta in 1 hr which reaches its peak at 24 hr on application of force thus supports the concept that inflammation plays a vital role during orthodontic tooth movement.

Interleukin-1 beta and root resorption:

The process of resorption was defined using various terminology namely apical root resorption (ARR), external apical root resorption (EARR), orthodontically induced root resorption (OIRR), orthodontically induced inflammatory root resorption (OIIRR) and others.^[51-55] Recently the new term orthodontitis is being introduced to define this process.^[56] Based on the radiographic findings orthodontitis can be divided into two groups as instrumental orthodontitis and instrumental – detrimental orthodontitis.^[57]

The inflammation process which is an important feature of tooth movement is also an important component behind root resorption process. Even though the cellular process underlying tooth movement and root resorption is thought to be similar evidence based data related to the molecular basis of root resorption is comparatively less at this point. Yamaguchi and Garlet^[2] considered the possibility of this similar inflammatory mechanism to be operated as a constructive one mediating the tooth movement and as destructive one that ends up in root resorption. He further added that there exists an unclarity about the driving factor which dictates constructive or destructive inflammation. Studies have reported the role of inflammatory mediators and chemokines in root resorption process.^[58-60] Diercke et al^[61] reported in his study the role IL-1 beta and a compressive force in inducing the production of RANKL in cementoblasts suggesting that particular cell type on activation could dictate root

resorption in the apex area. For a long time, it has been suggested to use lighter force to overcome root resorption. Later regime of force application was related to root resorption and studies have been conducted and recommended to use an intermittent type of force as it causes less severe root resorption than the continuous type of force.^[62] Root resorption is affected by both patient and treatment related factors. Studies done by Harris et al^[63] and Al-Qawasami et al^[64] can be considered as a milestone in relating genetics and root resorption. In 2003 Al-Qawasami et al^[64] is the first one to report the association of EARR and IL-1beta polymorphism thereby emphasizing the role of IL-1 beta gene polymorphism in external apical root resorption. They have also suggested to screen orthodontic patients for IL-1beta genotype and to identify those who carry 2 copies of allele 1 of IL-1beta by analysing the DNA which might help to identify people who are at risk of developing root resorption before orthodontic treatment. Following this, various others studies^[65-70] replicated the probable association of IL-1 beta gene polymorphism with root resorption and also explored other gene polymorphism such as vitamin D receptor.^[71] Gulden et al^[72] in his retrospective study found that IL-1 beta gene polymorphism does not predispose external apical root resorption. While another study^[66] found that not only variation in IL-1 beta but also variation in IL-1RN are responsible for post orthodontic external apical root resorption. Hope further research in future will resolve the debate over the role of IL-1 beta gene variation and other biological agents like interleukins, RANK and RANKL, prostaglandins and OPG in predisposing external apical root resorption. Iglesias-Linares and Hartsfield^[73] suggested that root resorption results not from a single pathway or factor but by the interaction of several pathways at the level of cell fusion/activation, clastic cell adhesion, and mineralized tissue reparative capabilities

Interleukin-1 beta and alveolar bone remodelling:

The osseous response that permits tooth movement in orthodontics results from functional load overlaid by the static load to produce a dynamic load. [74] The bone remodelling process during OTM is a result of effective interaction between osteoclasts, osteoblasts, osteocytes and PDL cells. When the mechanical load is applied during OTM the PDL mediated immune response induces the inflammatory response results in bone remodelling that involves many regulators. Cytokines are one among them. Cytokines possess not only osteoclastogenic activity but also anti osteoclastogenic properties and there by maintains the bone homeostasis. IL-1beta is a potent initiator of osteoclast differentiation. [75]

During initial stages of orthodontic tooth movement, IL-1beta is produced in the compression side of periodontal space and it has a direct effect on bone resorption. [76]

IL-1beta produced in response to orthodontic force stimulates osteoclastogenesis. [33,37] By induction of TNF- α and upregulation of RANKL & MMPs, IL1beta participate in several stages of osteoclastogenesis and determines the bone remodelling process. [75,77,78] The role of these factors in orthodontic tooth movement is well co-ordinated. [79] This phenomenon was confirmed in a mice model in which the administration of exogenous IL-1beta receptor antagonist results in not only a reduction in a number of osteoclasts but also the rate of tooth movement. [80] The inter-connection between the immune and skeletal system, the basis of osteoimmunology can be well explained using OTM model. Understanding this complex interaction in the orthodontic field is in its early stage and in future, works in this area will give us more in depth knowledge about osteoimmunology and tooth movement.

CONCLUSION

The production and activation of Interleukin 1 beta are certain during

orthodontic tooth movement as the underlying biological response is inflammatory in nature. Even though earlier attempts have been made to understand the influence of intensity of force on the expression level of IL-1 beta, the recent focus of research is on the effect of a regime of force application. IL-1 beta not only have an effect on bone remodelling and tooth movement but also on the various other associated phenomena such as pain, root resorption. There might be a cross talk between the pathways leading to bone remodelling, pain, tooth movement, and root resorption as they all have inflammation as a common thread. Current researches on biological response during tooth movement are focusing on a panel of associated or relevant biomarkers or genes instead of one. From this review it is obvious that when designing such a panel, IL-1 beta will definitely find a place in it.

REFERENCES

1. Chen. G.Y and Nunez. G. Sterile inflammation: sensing and reacting to damage. *Nat Rev Immunol.* 2010 December; 10(12): 826–837.
2. Yamaguchi M and Garlet G P. The role of inflammation in defining the type and pattern of tissue response in orthodontic tooth movement. In: *Biological Mechanisms of Tooth Movement*. Second edition. Edited by Vinod Krishnan and Ze'ev Davidovitch. published by John Wiley & Sons Ltd. 2015.pg 121-137.
3. David M Z. Genetic influence on orthodontic tooth movement. In: *Biological Mechanisms of Tooth Movement*. Second edition. Edited by Vinod Krishnan and Ze'ev Davidovitch. published by John Wiley & Sons Ltd. 2015.pg-147-163.
4. Alhashimi N, Frithiof L, Brudvik P et al., Orthodontic tooth movement and de novo synthesis of proinflammatory cytokines. *Am J Orthod Dentofacial Orthop.* 2001; 119:307–12.
5. Garlanda C, Dinarello CA, Mantovani A. The interleukin-1 family: back to the future. *Immunity.* 2013 Dec 12;39(6):1003-18.
6. Dinarello CA. IL-1: discoveries, controversies and future directions. *Eur J Immunol.* 2010 Mar;40(3):599-606.

7. Dinarello CA. A clinical perspective of IL-1 β as the gatekeeper of inflammation. *Eur J Immunol.* 2011 May;41(5):1203-17.
8. Bodel P, Miller H. Differences in pyrogen production by mononuclear phagocytes and by fibroblasts or HeLa cells. *J Exp Med.* 1977 Mar 1;145(3):607-17.
9. Bodel P, Ralph P, Wenc K et al., Endogenous pyrogen production by Hodgkin's disease and human histiocytic lymphoma cell lines in vitro. *J Clin Invest.* 1980 Feb;65(2):514-8.
10. Dinarello CA. Biologic basis for interleukin-1 in disease. *Blood.* 1996 Mar 15;87(6):2095-147.
11. Oliveira C M B, Sakata R K, Issy A Met al., Cytokines and Pain. *Rev Bras Anesthesiol* 2011;61(2): 255-265.
12. Long H, Wang Y, Jian F et al., Current advances in orthodontic pain. *Int J Oral Sci.* 2016 Jun 30;8(2):67-75.
13. Kobayashi M, Horinuki E. Neural mechanisms of nociception during orthodontic treatment. *J Oral Sci.* 2017; 59(2):167-171.
14. Coutaux A, Adam F, Willer JC et al., Hyperalgesia and allodynia: peripheral mechanisms. *Joint Bone Spine.* 2005 Oct;72(5):359-71.
15. Luppapanornlarp S, Kajii TS, Surarit R et al., Interleukin-1beta levels, pain intensity, and tooth movement using two different magnitudes of continuous orthodontic force. *Eur J Orthod.* 2010 Oct;32(5):596-601.
16. Davidovitch, Z., Nicolay, O, Alley, K et al, First and second messengers in interactions in stressed connective tissues in vitro. In: Norton L., Burstone C.J. (Eds.) *The Biology of Tooth Movement.* CRC Press, Boca Raton, FL; 1989:97-130.
17. Shavit Y, Fridel K, Beilin B. Postoperative pain management and proinflammatory cytokines: animal and human studies. *J Neuroimmune Pharmacol.* 2006 Dec;1(4):443-51.
18. Richardson JD, Vasko MR. Cellular mechanisms of neurogenic inflammation. *J Pharmacol Exp Ther.* 2002 Sep; 302(3):839-45.
19. Maggi CA. Tachykinins and calcitonin gene-related peptide (CGRP) as co-transmitters released from peripheral endings of sensory nerves. *Prog Neurobiol.* 1995 Jan;45(1):1-98.
20. Sommer C, Kress M. Recent findings on how proinflammatory cytokines cause pain: peripheral mechanisms in inflammatory and neuropathic hyperalgesia. *Neurosci Lett.* 2004 May 6;361(1-3):184-7.
21. Poole S, Cunha FQ, Ferreira SH: Hyperalgesia from subcutaneous cytokines. In: Watkins LR, Maier SF (eds): *Cytokines and Pain.* Basel: Birkhauser, 1999, pp 59-87
22. Fukuoka H, Kawatani M, Hisamitsu T et al., Cutaneous hyperalgesia induced by peripheral injection of interleukin-1 beta in the rat. *Brain Res.* 1994 Sep 19;657(1-2):133-40.
23. Yamaguchi M, Yoshii M, Kasai K. Relationship between substance P and interleukin-1beta in gingival crevicular fluid during orthodontic tooth movement in adults. *Eur J Orthod.* 2006;28(3):241-6.
24. Giannopoulou C, Dudic A, Kiliaridis S. Pain discomfort and crevicular fluid changes induced by orthodontic elastic separators in children. *J Pain.* 2006 May;7(5):367-76.
25. Ren Y, Vissink A. Cytokines in crevicular fluid and orthodontic tooth movement. *Eur J Oral Sci.* 2008 Apr;116(2):89-97.
26. Dudic A, Kiliaridis S, Mombelli A et al., Composition changes in gingival crevicular fluid during orthodontic tooth movement: comparisons between tension and compression sides. *Eur J Oral Sci.* 2006 Oct;114(5):416-22.
27. Hill PA. Bone remodelling. *Br J Orthod.* 1998 May;25(2):101-7.
28. Lekic P, McCulloch CA. Periodontal ligament cell population: the central role of fibroblasts in creating a unique tissue. *Anat Rec.* 1996 Jun;245(2):327-41.
29. Bourauel C, Vollmer D, Jäger A. Application of bone remodeling theories in the simulation of orthodontic tooth movements. *J Orofac Orthop.* 2000; 61(4):266-79.
30. Kawarizadeh A, Bourauel C, Jäger A. Experimental and numerical determination of initial tooth mobility and material properties of the periodontal ligament in rat molar specimens. *Eur J Orthod.* 2003 Dec;25(6):569-78.
31. Meager A. Cytokine regulation of cellular adhesion molecule expression in inflammation. *Cytokine Growth Factor Rev.* 1999 Mar;10(1):27-39.

32. Grieve WG 3rd, Johnson GK, Moore RN et al., Prostaglandin E (PGE) and interleukin-1 beta (IL-1 beta) levels in gingival crevicular fluid during human orthodontic tooth movement. *Am J Orthod Dentofacial Orthop.* 1994 Apr;105(4):369-74.
33. Uematsu S, Mogi M, Deguchi T. Interleukin (IL)-1 beta, IL-6, tumor necrosis factor-alpha, epidermal growth factor, and beta 2-microglobulin levels are elevated in gingival crevicular fluid during human orthodontic tooth movement. *J Dent Res.* 1996 Jan;75(1):562-7.
34. Wu CC, Li YS, Haga JH, et al. Roles of MAP kinases in the regulation of bone matrix gene expressions in human osteoblasts by oscillatory fluid flow. *J Cell Biochem.* 2006 Jun 1;98(3):632-41.
35. Cowin SC, Weinbaum S. Strain amplification in the bone mechanosensory system. *Am J Med Sci.* 1998 Sep;316(3):184-8.
36. Jendrucko RJ, Hyman WA, Newell PH Jr et al., Theoretical evidence for the generation of high pressure in bone cells. *J Biomech.* 1976;9(2):87-91.
37. Ren Y, Hazemeijer H, de Haan B et al., Cytokine profiles in crevicular fluid during orthodontic tooth movement of short and long durations. *J Periodontol.* 2007 Mar;78(3):453-8.
38. Kawasaki K, Takahashi T, Yamaguchi M et al., Effects of aging on RANKL and OPG levels in gingival crevicular fluid during orthodontic tooth movement. *Orthod Craniofac Res.* 2006 Aug;9(3):137-42.
39. Başaran G, Ozer T, Kaya FA et al., Interleukin-1 beta and tumor necrosis factor-alpha levels in the human gingival sulcus during orthodontic treatment. *Angle Orthod.* 2006 Sep;76(5):830-6.
40. Iwasaki LR, Crouch LD, Tutor A et al., Tooth movement and cytokines in gingival crevicular fluid and whole blood in growing and adult subjects. *Am J Orthod Dentofacial Orthop.* 2005 Oct;128(4):483-91.
41. Tzannetou S, Efstratiadis S, Nicolay O et al., Interleukin-1 beta and beta-glucuronidase in gingival crevicular fluid from molars during rapid palatal expansion. *Am J Orthod Dentofacial Orthop.* 1999 Jun;115(6):686-96.
42. Atuğ Özcan SS, Ceylan I, Ozcan E et al., Evaluation of oxidative stress biomarkers in patients with fixed orthodontic appliances. *Dis Markers.* 2014; 2014:1-7.
43. Ribagin LS, Rashkova MR. Matrix metalloproteinase-8 and interleukin-1 beta in gingival fluid of children in the first three months of orthodontic treatment with fixed appliances. *Folia Med (Plovdiv).* 2012 Jul-Sep;54(3):50-6.
44. Lee KJ, Park YC, Yu HS et al., Effects of continuous and interrupted orthodontic force on interleukin-1 beta and prostaglandin E2 production in gingival crevicular fluid. *Am J Orthod Dentofacial Orthop.* 2004 Feb;125(2):168-77.
45. Iwasaki LR, Gibson CS, Crouch LD et al., Speed of tooth movement is related to stress and IL-1 gene polymorphisms. *Am J Orthod Dentofacial Orthop.* 2006 Dec;130(6):698 e1-9.
46. Iwasaki LR, Haack JE, Nickel JC et al., Human interleukin-1 beta and interleukin-1 receptor antagonist secretion and velocity of tooth movement. *Arch Oral Biol.* 2001 Feb;46(2):185-9.
47. Iwasaki LR, Chandler JR, Marx DB et al., IL-1 gene polymorphisms, secretion in gingival crevicular fluid, and speed of human orthodontic tooth movement. *Orthod Craniofac Res.* 2009; 12(2):129-40.
48. Frodge BD, Ebersole JL, Kryscio RJ et al., Bone remodelling biomarkers of periodontal disease in saliva. *J Periodontol.* 2008 Oct;79(10):1913-9.
49. Saadi N, Ghaib N H. Effect of orthodontic tooth movement on salivary levels of Interleukin-1 beta, Tumor Necrosis Factor-alpha, and C-reactive protein. *J Bagh Coll Dentistry* 2013; 25(4):120-125.
50. Luppapornlarp S and Iida J. Orthodontic force, tooth movement, and interleukin-1 beta. *Hokkaido J. Dent. Sci.*, 38 (Special issue) : 20-27, 2017.
51. Linge BO, Linge L. Apical root resorption in upper anterior teeth. *Eur J Orthod.* 1983 Aug;5(3):173-83.
52. Parker RJ, Harris EF. Directions of orthodontic tooth movements associated with external apical root resorption of the maxillary central incisor. *Am J Orthod Dentofacial Orthop.* 1998 Dec;114(6):677-83.
53. Owman-Moll P, Kurol J, Lundgren D. Repair of orthodontically induced root resorption in adolescents. *Angle Orthod.* 1995;65(6):403-8.

54. Brezniak N, Wasserstein A. Orthodontically induced inflammatory root resorption. Part I: The basic science aspects. *Angle Orthod.* 2002 Apr;72(2):175-9.
55. Brezniak N, Wasserstein A. Orthodontically induced inflammatory root resorption. Part II: The clinical aspects. *Angle Orthod.* 2002 Apr;72(2):180-4.
56. Brezniak N, Wasserstein A. Defining and framing orthodontitis: a new term in orthodontics. *Angle Orthod.* 2014 May;84(3):568-9.
57. Brezniak N and Wasserstein A. Orthodontitis: The Inflammation Behind Tooth Movement and Orthodontic Root Resorption. In: *Biology of Orthodontic Tooth Movement-Current Concepts and Applications in Orthodontic Practice.* Edited by Bhavna Shroff. Springer International Publishing Switzerland 2016.pg-67-101
58. Curl L, Sampson W. The presence of TNF-alpha and TNFR1 in aseptic root resorption. A preliminary study. *Aust Orthod J.* 2011 Nov;27(2):102-9.
59. Asano M, Yamaguchi M, Nakajima R et al., IL-8 and MCP-1 induced by excessive orthodontic force mediates odontoclastogenesis in periodontal tissues. *Oral Dis.* 2011 Jul;17(5):489-98.
60. Yamaguchi M, Ukai T, Kaneko T et al., T cells are able to promote lipopolysaccharide-induced bone resorption in mice in the absence of B cells. *J Periodontal Res.* 2008 Oct;43(5):549-55.
61. Diercke K, Kohl A, Lux CJ et al., IL-1 β and compressive forces lead to a significant induction of RANKL-expression in primary human cementoblasts. *J Orofac Orthop.* 2012 Sep;73(5):397-412.
62. Acar A, Canyürek U, Kocaaga M et al., Continuous vs. discontinuous force application and root resorption. *Angle Orthod.* 1999 Apr;69(2):159-63.
63. Harris EF, Kineret SE, Tolley EA. A heritable component for external apical root resorption in patients treated orthodontically. *Am J Orthod Dentofacial Orthop.* 1997 Mar;111(3):301-9.
64. Al-Qawasmi RA, Hartsfield JK Jr, Everett ET et al., Genetic predisposition to external apical root resorption. *Am J Orthod Dentofacial Orthop.* 2003 Mar;123(3):242-52.
65. Bastos Lages EM, Drummond AF, Pretti H et al., Association of functional gene polymorphism IL-1beta in patients with external apical root resorption. *Am J Orthod Dentofacial Orthop.* 2009 Oct;136(4):542-6.
66. Iglesias-Linares A, Yañez-Vico R, Ballesta-Mudarra S et al., Postorthodontic external root resorption is associated with IL1 receptor antagonist gene variations. *Oral Dis.* 2012 Mar;18(2):198-205.
67. Linhartova P, Cernochova P, Izakovicova Holla L. IL1 gene polymorphisms in relation to external apical root resorption concurrent with orthodontia. *Oral Dis.* 2013 Apr;19(3):262-70.
68. Pereira S, Lavado N, Nogueira L, et al. Polymorphisms of genes encoding P2X7R, IL-1B, OPG and RANK in orthodontic-induced apical root resorption. *Oral Dis.* 2014;20:659–667.
69. Rossi M, Whitcomb S, Lindemann R. Interleukin-1 beta and tumor necrosis factor-alpha production by human monocytes cultured with L-thyroxine and thyrocalcitonin: relation to severe root shortening. *Am J Orthod Dentofacial Orthop.* 1996 Oct;110(4):399-404.
70. Tomoyasu Y, Yamaguchi T, Tajima A, et al., External apical root resorption and the interleukin-1B gene polymorphism in the Japanese population. *Orthodontic waves.* 2009;68;pg-152–157.
71. Fontana ML, de Souza CM, Bernardino JF et al., Association analysis of clinical aspects and vitamin D receptor gene polymorphism with external apical root resorption in orthodontic patients. *Am J Orthod Dentofacial Orthop.* 2012 Sep;142(3):339-47.
72. Gülden N, Eggermann T, Zerres K et al, Interleukin-1 polymorphisms in relation to external apical root resorption (EARR). *J Orofac Orthop.* 2009 Jan;70(1):20-38.
73. Iglesias-Linares A, Hartsfield JK Jr. Cellular and Molecular Pathways Leading to External Root Resorption. *J Dent Res.* 2017 Feb;96(2):145-152.
74. Roberts WE. Rigid endosseous anchorage and tricalcium phosphate (TCP)-coated implants. *CDA J.* 1984 Dec;12(12):158-61.
75. Ruscitti P, Cipriani P, Carubbi F et al., The role of IL-1 β in the bone loss during rheumatic diseases. *Mediators Inflamm* 2015; vol. 2015, Article ID 782382, 10 pages.
76. Bletsa A, Berggreen E, Brudvik P. Interleukin-1alpha and tumor necrosis

- factor-alpha expression during the early phases of orthodontic tooth movement in rats. *Eur J Oral Sci.* 2006 Oct;114(5):423-9.
77. Teixeira CC, Khoo E, Tran J et al., Cytokine expression and accelerated tooth movement. *JDent Res.* 2010 Oct;89(10):1135-41.
78. Wei S, Kitaura H, Zhou P et al., IL-1 mediates TNF-induced osteoclastogenesis. *J Clin Invest.* 2005 Feb;115(2):282-90.
79. Yamaguchi M, Kasai K. Inflammation in periodontal tissues in response to mechanical forces. *Arch Immunol Ther Exp (Warsz).* 2005 Sep-Oct;53(5):388-98.
80. Lee TY, Lee KJ, Baik HS. Expression of IL-1beta, MMP-9 and TIMP-1 on the pressure side of gingiva under orthodontic loading. *Angle Orthod.* 2009 Jul;79(4):733-9.

How to cite this article: Lalithapriya S, Rajasigamani K, Bhaskar V. Role of interleukin-1 beta in orthodontics. *Int J Health Sci Res.* 2018; 8(11):270-278.
