

Original Research Article

Bacteremia after Dental Procedures

Sunita Bhatawadekar¹, Renu Bhardwaj²

¹Department of Microbiology, Bharati Vidyapeeth Deemed University Medical College, Pune. ²Department of Microbiology, B.J. Medical College, Pune, India.

Corresponding Author: Sunita Bhatawadekar

Received: 27/11/2014

Revised: 16/12/2014

Accepted: 22/12/2014

ABSTRACT

Aims and objective: Dental extraction and scaling are very simple dental procedures carried out routinely in most outpatient departments. However, these seemingly innocuous procedures have been known to result in transient bacteremia which may cause problems in patients with cardiac valve defects, with prosthetic joints, in patients with renal transplant and in immunocompromised patients. In pregnant women with periodontal disease transient bacteremia increases three to five times greater risk of preterm birth. In the present study, the incidence, type, intensity and duration of bacteremia following dental procedure was assessed.

Materials and methods: Aerobic and anaerobic blood culture and quantitative estimation of bacteremia was studied for up to 30 min. after dental procedures. As a control group bacteremia was assessed in ten individuals after mastication.

Results: The incidence of bacteremia was 80% after dental procedure. In 50 % patients bacteremia was observed at 10 min. after the procedure and in 26 % it persisted for 30 min. More than 30 CFU /ml of blood were present in 6% patients at 10 min. and in 4% patients at 30 min. A high incidence of bacteremia was reported after dental extraction (88%), after dental scaling (75%) and after intra oral wire removal (100%) Aerobes were isolated from 73.33% of the blood cultures. Common aerobes isolated were alpha hemolytic streptococci and Actinomyces spp. Anaerobes were isolated from 26.66% of the blood cultures. Common anaerobes encountered were Prevotella spp. and Peptostreptococci spp.

Conclusions: Since bacteremia following dental procedures is poly microbial and usually caused by oral flora, adequate prophylaxis should be directed at decreasing the oral flora by application of local antiseptics and chemo prophylaxis with broad spectrum antibiotics prior to every minor dental surgical procedure.

Key words – Transient bacteremia, dental procedures, blood culture.

INTRODUCTION

Modern dietary habits have increased the incidence of dental problems in today's society. Caries and periodontitis continue to be major public health problems throughout the world. Many dental procedures are carried out routinely as outpatient procedures in the OPD. However, these seemingly innocuous procedures may occasionally be associated with serious complications.

Dental procedures on patients with gingivitis and periodontitis are likely to result in transient bacteremia. ^[1] A nidus of these organisms settles at various sites during the phase of bacteremia and later multiply leading to life threatening complications. This bacteremia may result in bacterial endocarditis in patients with cardiac valve defects.^[2] Patients having prosthetic joint replacement may lose a prosthetic joint because of bacteremia following dental procedures ^[3] In patients on haemodialysis or with kidney transplant, it may result in renal infection. Lastly with the increasing incidence of HIV, it may lead deep-seated abscesses in an immunocompromised patient. In pregnant women with periodontal disease, transient bacteremia increases three to five times greater risk of preterm birth.^[5]

In the present study, incidence, duration, intensity of bacteremia following dental bacteremia was assessed and efforts were made to find out the common causative agents of bacteremia.

MATERIALS AND METHODS

Fifty patients undergoing various dental procedures were studied. Informed consent was taken from all the patients. All the patients in this study group were given Cotrimazzole for three days prior to the procedures. All the dental procedures were carried out under local anesthesia. 10 ml of blood was collected ten minutes and thirty after dental procedure minutes and processed as follows.4 ml of blood was used to quantitate the bacteremia by pour plate technique.3 ml of blood was added to 30 ml of freshly prepared thioglycolate broth containing resazurine as an indicator; this broth was used to cultivate microaerophilic After and the anaerobic organisms. inoculating blood in to the broth air was evacuated from the bottle and bottle flushed with carbon dioxide. 3 ml of blood was

added to the 30 ml of tryptose phosphate broth for aerobic blood culture.

The thioglycolate broth was plated on Neomycin blood agar and Kanamycin vancomycin blood agar. All strict anaerobes were subjected to biochemical test and the isolates were identified according to the Wadsworth anaerobic manule.^[6]

For aerobic organisms subcultures were made on 2nd, 4th and 7th day. The media inoculated were blood agar and Mac conkey's agar. Isolates were identified according to Mackie and Macartney.^[7]

A control group of ten individuals with good dental hygiene were also included in the study. Each individual was asked to chew sugarcane for five minutes. Blood samples were collected at 10 minutes and 30 minutes after mastication. These were processed in the same manner as the test group.

OBSERVATIONS AND RESULTS

Incidence of bacteremia in the test group was 80% and in control group it was 20%. The incidence of bacteremia was significant in test group using the Fisher exact test.

 Table 1. Incidence of bacteremia in relation to dental procedures in test group

proce	procedures in test group				
No	Dental procedures	No. Of patients with bacteremia	Percentage		
1	Extraction (27)	24	48		
2	Scaling (8)	06	12		
3	Tooth rocking (5)	03	06		
4	Wire removal (3)	03	06		
5	Removal of bony spicules (2)	02	04		
6	Dental filling (3)	01	02		
7	Root canal (1)	01	02		
8	Excision of epulis (1)	00	00		
	Total	40	80		

(Table 1) In the present study the incidence of bacteremia following various dental procedures was 80%. The incidence of bacteremia was maximum after more traumatic procedures such as wire removal

and extraction. It was minimum for minor dental procedures such as dental fillings.

Table. 2- Incidence of bacteremia following dental procedures with relation to time interval for collection of blood.

** 1011 1	the relation to three meet varior concetion of blood.				
Sr	Time interval for	No. of patients	percentage		
no.	collection of blood sample	with bacteremia			
1	Only 10 min positive	25	50		
2	10 min and 30 min positive	13	26		
3	10 min negative and 30 min positive	02	04		
4	10 min and 30 min negative	10	20		

Table 3- Quantitative study of 10 minutes and 30 minutes blood samples indicating intensity of aerobic and anaerobic bacteremia following dental procedures

Colony	10 minutes		30 minutes	
forming units	forming units			
/ml of blood				
	No of patients	Percentage	No of patients	Percentage
1-5	33	66	10	20
6-10	04	08		
11 - 20				
21 - 30			01	02
More than 30	03	06	02	04
No growth	10	20	37	74

Table. 4- Correlation of degree of aerobic and anaerobic bacteremia with relation to time period after the dental procedure.

	Type of incubation	Patients with quantifiable	
		organisms	
		No.	Percentage
10 minutes	Aerobic	24	60
	Anaerobic	16	40
	Total	40	100
30 minutes	Aerobic	07	53.84
	Anaerobic	06	46.16
	Total	13	100

(Table 2) In 50% patients bacteremia was observed after 10 min of dental procedures. In 26% of patients it persisted for 30 minutes. In 4% patients late onset bacteremia was observed and bacteria were isolated only from 30 minutes blood samples. (Table 3) Intensity of bacteremia was low. (Table 4)Out of forty patients with bacteremia, in 24 patients aerobic organisms were present at 10 minutes. However, they persisted for up to 30 minutes after dental procedures, in only 7 patients. In 16 patients anaerobic organisms were present at 10 minutes and in 6 patients persisted up to 30 minutes, after dental procedures. Anaerobes seem to persist in the blood for a longer time. Aerobic and anaerobic mixed type of bacteremia was present in many patients.

Table -5 Types of aerobic and facultatively anaerobic microorganisms isolated from blood cultures after dental procedures.

procedu	procedures.				
Sr. no	Type of organism	No of patients	Percentage		
1	Streptococci	25	37.87		
2	Actinomyces spp.	12	18.17		
3	Diptheroids	11	16.66		
4	Staphylococci	08	12.11		
5	Campylobacte spp.	03	04.54		
6	Bacillus spp.	02	03.03		
7	Citrobacter spp	01	01.51		
8	Candida spp.	01	01.51		
9	Haemophilus spp.	01	01.51		
10	Moraxilla spp.	01	01.51		
11	Unindentified Gram	01	01.51		
	negative coccobacilla				
	Total	66	100		

Table 6. Type of anaerobic organisms isolated from blood cultures after dental procedures.

Sr no	Type of organisms	No of	Percentage
		patients	
1	Peptrostreptocopccus spp.	06	25.00
2	Prevotella melaninogenicus	04	16.66
3	Unidentified Gram positive	04	16.66
	bacilli		
4	Bacteroides spp.	04	16.66
5	Peptococcus spp	02	8.33
6	Veillonella parvula	02	8.33
7	Fusobacterium nucleatum	01	4.16
8	Clostridium spp	01	4.16
	Total	24	100

(Table 5 and 6). Alpha hemolytic Streptococci, Actinomyces, Peptostreptococci Prevotella species. species and Bacteroides species were most commonly isolated from blood cultures following dental manipulations. Alpha hemolytic Streptococcal species isolated were S.mitis, S.intermedius, S, salivarious, S, sangiis. Variety of Actinomyces isolated from blood samples after dental procedures A.viscosus. A, odontolyticus, were A.naeslundii.

DISCUSSION

Blood is usually sterile, but transient bacteremia can occur after manipulations of infective focus.^[8,9] Every dental procedure,

even minor dental manipulation may result in transient bacteremia.

In the present study overall incidence of bacteremia was 80%. Various workers have reported an incidence of bacteremia ranging from 15 to 100%. ^[10-14] In Indian patients incidence of bacteremia was 52% after dental procedures. (Pol D G. Unpublished data 1981, dissertation.).

Incidence of bacteremia following dental procedures was maximum after traumatic procedures and procedures involving handling of periodontal tissue. (Table 1) It was minimum for minor dental procedures such as dental fillings. Similar results has been reported by Rogosa et al.^[12]

In the present study intensity of bacteremia was low. Similar low intensity has been reported by other workers also. ^[1,14] (Table 3)

Ninety different types of organisms were isolated from forty bacteremic patients. Out of these, 66 (77.33%) were aerobes and facultative anaerobes, where as 24 (26. 66 %) were obligatory anaerobes. Alpha hemolytic Streptococci and Actinomyces spp. were most commonly isolated from blood cultures after dental procedures.

Streptococci comparable to the present Study were reported by various workers. ^[15,16] Actinomyces strains are regular inhabitants of the oral cavity and play an important role in periodontitis. Actinomyces have been isolated from Indian patients suffering with gingivitis and periodontitis. ^[17]

Actinomyces causing bacteremia following dental procedures has been reported to range from 8. 54% to 14.73%. [15,16] Peptostreptococci (33.32%), Prevotella spp. (16. 66%) and Bacteroides spp (16.66%) were commonly isolated from blood cultures after dental procedures.

Dental manipulations leading to bacteremia can act as a potential focus of endogenous infection. Various infections linked to the oral cavity, at present reported in literature may be representing only the tip of the iceberg. Increased awareness may generate evidence linking these diseases to bacteremia resulting from dental manipulation.

Bacteremia after dental manipulations may have long-term sequlae. So frequent follow up and prophylactic antibiotics for patients undergoing dental manipulations are required. According to American Heart Association recommendations only people with highest risk of developing infective endocarditis should receive antibiotics.

Antibiotic prophylaxis is not generally recommended for people with moderate risk conditions.^[18]

When treating patients with heart conditions, dentists follow recommendations developed by the American Heart Association (AHA), with input from the American Dental Association. For patients who have total replacements, joint they refer to recommendations developed by the American Academy of Orthopaedic Surgeons (AAOS).

The best approach to control this bacteremia would be preoperative preparation of the oral cavity with a view to decrease the bacterial load at the time of dental procedure. Important dental care recommendations are -Anyone who is at risk of developing endocarditis should follow a program of careful mouth and tooth care. This includes a professional cleaning every six months, twice daily tooth brushing, and once daily flossing. These measures can help to prevent plaque and bacteria from building up around the gums and teeth.

REFERENCES

1. Okell C C, Elliot S D, Bacteremia and oral sepsis. Lancet 1935; 2: 869-2.

- Kaplin E K.Anderson R C. Infective endocarditis after use of dental irrigation devise. Lancet 1977; 2:610
- Thyne GM, Ferguson J W, Antibiotic prophylaxis during dental treatment in patients with prosthetic joints. J. Bone Joint surg. 1991; 73B(2): 191-4.
- 4. Lucartorto F. M, Colin K. F. Maza. J., Post scaling bacteremia in HIV associated gingivitis and periodontitis. Oral Surg Oral Med Oral Pathol. 1992; 73:550-4.
- Ananda P. Dasanayake, Yihong Li, Howard Wiener, John D. Ruby, Men-Jean Lee.Salivary Actinomyces naeslundii Genospecies 2 and Lactobacillus casei Levels Predict Pregnancy Outcomes. J Periodontol February 2005, Vol. 76, No. 2, Pages 171-177.
- Sutter V.L, Citron D. M., Edelstein MAC, Finegold SM. Wadsworth anaerobic bacteriology manual. 4th Ed. Baltimore California. Star publishing company 1986.
- Colle J C, Dugid J P, Fraser A G, Marmion B P(Ed) Mackie and MaCartney's practical Medical Microbiology vol. 2, 13th Ed, Edinburg Churchill Livingstone 1989.
- Barrington F T, Wright H D. Bacteremia following operations on the urethra. J Pathl Bacterol. 1930; 33: 871-9.
- Southworth H, Flake C G. Blood culture after tonsillectomy. American Journal of Medical sciences. 1938; 195:667-670.
- Coffin E, Thompson REM Factors influencing bacteremia following dental extraction. Lancet.1956; 271: 654-6

- Schirger A, Martin W J, Royer Q, Needham G M. Bactereial invasion of blood after oral surgical procedures. J Lab Clin Med. 1960; 55: 376-380.
- 12. Rogosa M, Hampp E G, Nevin T A, et al. Blood sampling and cultural studies in the detection of postoperative bacteremias. J Am Dent Asso. 1960; 60: 171-180.
- 13. Craford J, Sconyers J R, Moriarthy J D, King R C. West J E. Bacteremia after tooth extractions studied with aid of prereduced anaerobically sterilised culture media. Applied Microbiology 1974; 27: 927-932.
- Heimdahl A, Gunner H, Hedberg et al. Detection and quantitation by lysis filtration of bacteremia after different oral surgical procedures. J Clin Microbiol. 1990; 28: 2205-9.
- 15. Khairat O. The nonaerobes of post extraction bacteremia. J Dent Res 1966; 45 (4): 1191-1197.
- 16. Sweet J B, Gill V J, Chusid M J, Elin R J. Nitroblue tetrazolium and limulus assays for bacteremia after dental extracts, effect of topical antiseptics. J Am Den Asso. 1978; 96: 276-281.
- 17. Saini S, Aparna, Gupta N, Mahajan A, Saini O. P. Antibiotic susceptibility of bacterial isolates in gingivitis and periodontitis. Indian Journal of Dental Research .2003; 14(2): 95-100.
- 18. Nishimura RA, Carabello BA, Faxon DP, et al. ACC/AHA 2008 guideline update on valvular heart disease: focused update infective on endocarditis: of a report the American College of Cardiology/American Heart Association Task Force on Practice Guidelines: endorsed by the Society of Cardiovascular Anesthesiologists,

Society for Cardiovascular Angiography and Interventions, and

Society of Thoracic Surgeons. Circulation 2008; 118:833.

How to cite this article: Bhatawadekar S, Bhardwaj R. Bacteremia after dental procedures. Int J Health Sci Res. 2015;5(1):113-118.

International Journal of Health Sciences & Research (IJHSR)

Publish your work in this journal

The International Journal of Health Sciences & Research is a multidisciplinary indexed open access double-blind peerreviewed international journal that publishes original research articles from all areas of health sciences and allied branches. This monthly journal is characterised by rapid publication of reviews, original research and case reports across all the fields of health sciences. The details of journal are available on its official website (www.ijhsr.org).

Submit your manuscript by email: editor.ijhsr@gmail.com OR editor.ijhsr@yahoo.com